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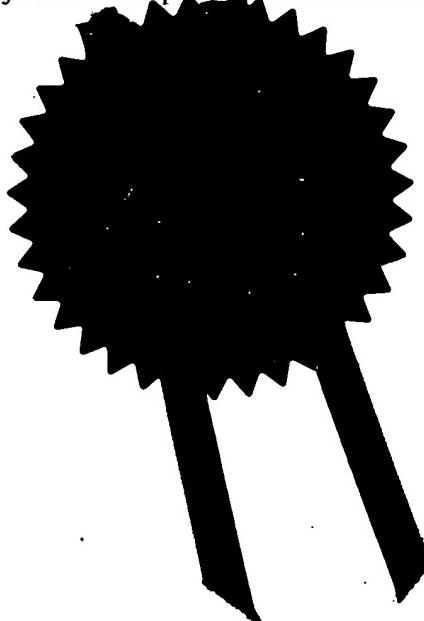
CERTIFIED COPY OF PRIORITY DOCUMENT

I, the undersigned, being an officer duly authorised in accordance with Section 74(1) and (4) of the Deregulation and Contracting Out Act 1994, to sign and issue certificates on behalf of the Comptroller-General, hereby certify that annexed hereto is a true copy of the international application filed on 22 February 2000 under the Patent Cooperation Treaty at the UK Receiving Office. The application was allocated the number PCT/GB2000/00624.

In accordance with the Patents (Companies Re-registration) Rules 1982, if a company named in this certificate and any accompanying documents has re-registered under the Companies Act 1980 with the same name as that with which it was registered immediately before re-registration save for the substitution as, or the inclusion as, the last part of the name of the words "public limited company" or their equivalents in Welsh, references to the name of the company in this certificate and any accompanying documents shall be treated as references to the name with which it is so re-registered.

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Signed 

Dated 04 August 2003

Home
Co

PCT

REQUEST

The undersigned requests that the present international application be processed according to the Patent Cooperation Treaty.

For receiving Office use only

B
PCT/GB 00/00624

International Application No.

22 FEBRUARY 2000

22.02.2000

International Filing Date

United Kingdom Patent Office
PCT International Application
Name of receiving Office and "PCT International Application"

Applicant's or agent's file reference
(if desired) (12 characters maximum) GWS/DC/21091

Box No. I TITLE OF INVENTION

NEISERIAL VACCINE COMPOSITIONS AND METHODS

Box No. II APPLICANT

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.)

MICROBIOLOGICAL RESEARCH AUTHORITY

CAMR

PORTON DOWN

SALISBURY, WILTSHIRE

SP4 OJG

GB

This person is also inventor.

Telephone No.

Faximile No.

Teleprinter No.

State (that is, country) of nationality:

GB

State (that is, country) of residence:

GB

This person is applicant all designated States all designated States except the United States of America the United States of America only the States indicated in the Supplemental Box

Box No. III FURTHER APPLICANT(S) AND/OR (FURTHER) INVENTOR(S)

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.)

ROBINSON, Andrew

MICROBIOLOGICAL RESEARCH AUTHORITY

CAMR

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SP4 OJG, GB

This person is:

applicant only

applicant and inventor

inventor only (If this check-box is marked, do not fill in below.)

State (that is, country) of nationality:

GB

State (that is, country) of residence:

GB

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Further applicants and/or (further) inventors are indicated on a continuation sheet.

Box No. IV AGENT OR COMMON REPRESENTATIVE; OR ADDRESS FOR CORRESPONDENCE

The person identified below is hereby/has been appointed to act on behalf of the applicant(s) before the competent International Authorities as:

agent

common representative

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.)

SCHLICH, George William

MATHYS & SQUIRE

100 Gray's Inn Road

London

WC1X 8AL

GB

Telephone No.

+44 (0)20 7830 0000

Faximile No.

+44 (0)20 7830 0001

Teleprinter No.

Address for correspondence: Mark this check-box where no agent or common representative is/has been appointed and the space above is used instead to indicate a special address to which correspondence should be sent.

Continuation of Box No. III

FURTHER APPLICANT(S) AND/OR (FURTHER) INVENTOR(S)

If none of the following sub-boxes is used, this sheet should not be included in the request.

(Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.)

GORRINGE, Andrew Richard

(c/o) MICROBIOLOGICAL RESEARCH AUTHORITY

CAMR

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SALISBURY, WILTSHIRE

SP4 OJG, GB

(If this check-box is marked, do not fill in below.)

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(Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.)

HUDSON, Michael John

(c/o) MICROBIOLOGICAL RESEARCH AUTHORITY

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BRACEGIRDLE, Philippa

(c/o) MICROBIOLOGICAL RESEARCH AUTHORITY

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SALISBURY, WILTSHIRE

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(Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.)

KROLL, John Simon

(c/o) IMPERIAL COLLEGE SCHOOL OF MEDICINE

ST MARY'S HOSPITAL

NORFOLK PLACE

LONDON W2 1PG

GB

(If this check-box is marked, do not fill in below.)

(that is, country)

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(that is, country)

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Continuation of Box No. III FURTHER APPLICANT(S) AND/OR (FURTHER) INVENTOR(S)

If none of the following sub-boxes is used, this sheet should not be included in the request.

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.)

CARTWRIGHT, Keith
Lero PUBLIC HEALTH LABORATORY SERVICE
HEADQUARTERS OFFICE
61 COLINDALE AVENUE
LONDON NW9 5DF
GB

This person is:

- applicant only
 applicant and inventor
 inventor only (If this check-box is marked, do not fill in below.)

State (that is, country) of nationality:
GBState (that is, country) of residence:
GBThis person is applicant
for the purposes of: all designated
States all designated States except
the United States of America the United States
of America only the States indicated in
the Supplemental Box

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.)

IMPERIAL COLLEGE SCHOOL OF SCIENCE, TECHNOLOGY AND
MEDICINE
SHERFIELD BUILDING
EXHIBITION ROAD
LONDON SW7 2AZ
GB

This person is:

- applicant only
 applicant and inventor
 inventor only (If this check-box is marked, do not fill in below.)

State (that is, country) of nationality:
GBState (that is, country) of residence:
GBThis person is applicant
for the purposes of: all designated
States all designated States except
the United States of America the United States
of America only the States indicated in
the Supplemental Box

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.)

PHLSB Public Health Laboratory Service Board
61 COLINDALE AVENUE
LONDON
NW9 5DF
GB

18306 17-5-2000

This person is:

- applicant only
 applicant and inventor
 inventor only (If this check-box is marked, do not fill in below.)

State (that is, country) of nationality:
GBState (that is, country) of residence:
GBThis person is applicant
for the purposes of: all designated
States all designated States except
the United States of America the United States
of America only the States indicated in
the Supplemental Box

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.)

State (that is, country) of nationality:

State (that is, country) of residence:

This person is applicant
for the purposes of: all designated
States all designated States except
the United States of America the United States
of America only the States indicated in
the Supplemental Box Further applicants and/or (further) inventors are indicated on another continuation sheet.

B. No.V DESIGNATION OF STATES

The following designations are hereby made under Rule 4.9(a) (mark the applicable check-boxes; at least one must be marked):

Regional Patent

- ARIPO Patent: GH Ghana, GM Gambia, KE Kenya, LS Lesotho, MW Malawi, SD Sudan, SL Sierra Leone, SZ Swaziland, TZ United Republic of Tanzania, UG Uganda, ZW Zimbabwe, and any other State which is a Contracting State of the Harare Protocol and of the PCT
- EA Eurasian Patent: AM Armenia, AZ Azerbaijan, BY Belarus, KG Kyrgyzstan, KZ Kazakhstan, MD Republic of Moldova, RU Russian Federation, TJ Tajikistan, TM Turkmenistan, and any other State which is a Contracting State of the Eurasian Patent Convention and of the PCT
- EP European Patent: AT Austria, BE Belgium, CH and LI Switzerland and Liechtenstein, CY Cyprus, DE Germany, DK Denmark, ES Spain, FI Finland, FR France, GB United Kingdom, GR Greece, IE Ireland, IT Italy, LU Luxembourg, MC Monaco, NL Netherlands, PT Portugal, SE Sweden, and any other State which is a Contracting State of the European Patent Convention and of the PCT
- OA OAPI Patent: BF Burkina Faso, BJ Benin, CF Central African Republic, CG Congo, CI Côte d'Ivoire, CM Cameroon, GA Gabon, GN Guinea, GW Guinea-Bissau, ML Mali, MR Mauritania, NE Niger, SN Senegal, TD Chad, TG Togo, and any other State which is a member State of OAPI and a Contracting State of the PCT (if other kind of protection or treatment desired, specify on dotted line)

National Patent (if other kind of protection or treatment desired, specify on dotted line):

- | | |
|--|--|
| <input checked="" type="checkbox"/> AE United Arab Emirates | <input checked="" type="checkbox"/> LR Liberia |
| <input checked="" type="checkbox"/> AL Albania | <input checked="" type="checkbox"/> LS Lesotho |
| <input checked="" type="checkbox"/> AM Armenia | <input checked="" type="checkbox"/> LT Lithuania |
| <input checked="" type="checkbox"/> AT Austria | <input checked="" type="checkbox"/> LU Luxembourg |
| <input checked="" type="checkbox"/> AU Australia | <input checked="" type="checkbox"/> LV Latvia |
| <input checked="" type="checkbox"/> AZ Azerbaijan | <input checked="" type="checkbox"/> MA Morocco |
| <input checked="" type="checkbox"/> BA Bosnia and Herzegovina | <input checked="" type="checkbox"/> MD Republic of Moldova |
| <input checked="" type="checkbox"/> BB Barbados | <input checked="" type="checkbox"/> MG Madagascar |
| <input checked="" type="checkbox"/> BG Bulgaria | <input checked="" type="checkbox"/> MK The former Yugoslav Republic of Macedonia |
| <input checked="" type="checkbox"/> BR Brazil | <input checked="" type="checkbox"/> MN Mongolia |
| <input checked="" type="checkbox"/> BY Belarus | <input checked="" type="checkbox"/> MW Malawi |
| <input checked="" type="checkbox"/> CA Canada | <input checked="" type="checkbox"/> MX Mexico |
| <input checked="" type="checkbox"/> CH and LI Switzerland and Liechtenstein | <input checked="" type="checkbox"/> NO Norway |
| <input checked="" type="checkbox"/> CN China | <input checked="" type="checkbox"/> NZ New Zealand |
| <input checked="" type="checkbox"/> CR Costa Rica | <input checked="" type="checkbox"/> PL Poland |
| <input checked="" type="checkbox"/> CU Cuba | <input checked="" type="checkbox"/> PT Portugal |
| <input checked="" type="checkbox"/> CZ Czech Republic | <input checked="" type="checkbox"/> RO Romania |
| <input checked="" type="checkbox"/> DE Germany | <input checked="" type="checkbox"/> RU Russian Federation |
| <input checked="" type="checkbox"/> DK Denmark | <input checked="" type="checkbox"/> SD Sudan |
| <input checked="" type="checkbox"/> DM Dominica | <input checked="" type="checkbox"/> SE Sweden |
| <input checked="" type="checkbox"/> EE Estonia | <input checked="" type="checkbox"/> SG Singapore |
| <input checked="" type="checkbox"/> ES Spain | <input checked="" type="checkbox"/> SI Slovenia |
| <input checked="" type="checkbox"/> FI Finland | <input checked="" type="checkbox"/> SK Slovakia |
| <input checked="" type="checkbox"/> GB United Kingdom | <input checked="" type="checkbox"/> SL Sierra Leone |
| <input checked="" type="checkbox"/> GD Grenada | <input checked="" type="checkbox"/> TJ Tajikistan |
| <input checked="" type="checkbox"/> GE Georgia | <input checked="" type="checkbox"/> TM Turkmenistan |
| <input checked="" type="checkbox"/> GH Ghana | <input checked="" type="checkbox"/> TR Turkey |
| <input checked="" type="checkbox"/> GM Gambia | <input checked="" type="checkbox"/> TT Trinidad and Tobago |
| <input checked="" type="checkbox"/> HR Croatia | <input checked="" type="checkbox"/> TZ United Republic of Tanzania |
| <input checked="" type="checkbox"/> HU Hungary | <input checked="" type="checkbox"/> UA Ukraine |
| <input checked="" type="checkbox"/> ID Indonesia | <input checked="" type="checkbox"/> UG Uganda |
| <input checked="" type="checkbox"/> IL Israel | <input checked="" type="checkbox"/> US United States of America |
| <input checked="" type="checkbox"/> IN India | <input checked="" type="checkbox"/> UZ Uzbekistan |
| <input checked="" type="checkbox"/> IS Iceland | <input checked="" type="checkbox"/> VN Viet Nam |
| <input checked="" type="checkbox"/> JP Japan | <input checked="" type="checkbox"/> YU Yugoslavia |
| <input checked="" type="checkbox"/> KE Kenya | <input checked="" type="checkbox"/> ZA South Africa |
| <input checked="" type="checkbox"/> KG Kyrgyzstan | <input checked="" type="checkbox"/> ZW Zimbabwe |
| <input checked="" type="checkbox"/> KP Democratic People's Republic of Korea | |
| <input checked="" type="checkbox"/> KR Republic of Korea | |
| <input checked="" type="checkbox"/> KZ Kazakhstan | |
| <input checked="" type="checkbox"/> LC Saint Lucia | |
| <input checked="" type="checkbox"/> LK Sri Lanka | |

Check-boxes reserved for designating States which have become party to the PCT after issuance of this sheet:

+ ANY OTHER STATE THAT IS PARTY TO THE PCT

Precautionary Designation Statement: In addition to the designations made above, the applicant also makes under Rule 4.9(b) all other designations which would be permitted under the PCT except any designation(s) indicated in the Supplemental Box as being excluded from the scope of this statement. The applicant declares that those additional designations are subject to confirmation and that any designation which is not confirmed before the expiration of 15 months from the priority date is to be regarded as withdrawn by the applicant at the expiration of that time limit. (Confirmation (including fees) must reach the receiving Office within the 15-month time limit.)

See Notes to the request form

Supplemental Box

If the Supplemental Box is not used, this sheet should not be included in the request.

1. If, in any of the Boxes, the space is insufficient to furnish all the information: in such case, write "Continuation of Box No." [indicate the number of the Box] and furnish the information in the same manner as required according to the captions of the Box in which space was insufficient, in particular:

- (i) if more than two persons are involved as applicants and/or inventors and no "continuation sheet" is available; in such case, write "Continuation of Box No. III" and indicate for each additional person the same type of information as required in Box No. III. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below;
- (ii) if, in Box No. II or in any of the sub-boxes of Box No. III, the indication "the States indicated in the Supplemental Box" is checked: in such case, write "Continuation of Box No. II" or "Continuation of Box No. III" or "Continuation of Boxes No. II and No. III" (as the case may be), indicate the name of the applicant(s) involved and, next to (each) such name, the State(s) (and/or, where applicable, ARIPO, Eurasian, European or OAPI patent) for the purposes of which the named person is applicant;
- (iii) if, in Box No. II or in any of the sub-boxes of Box No. III, the inventor or the inventor/applicant is not inventor for the purposes of all designated States or for the purposes of the United States of America: in such case, write "Continuation of Box No. II" or "Continuation of Box No. III" or "Continuation of Boxes No. II and No. III" (as the case may be), indicate the name of the inventor(s) and, next to (each) such name, the State(s) (and/or, where applicable, ARIPO, Eurasian, European or OAPI patent) for the purposes of which the named person is inventor;
- (iv) if, in addition to the agent(s) indicated in Box No. IV, there are further agents: in such case, write "Continuation of Box No. IV" and indicate for each further agent the same type of information as required in Box No. IV;
- (v) if, in Box No. V, the name of any State (or OAPI) is accompanied by the indication "patent of addition," or "certificate of addition," or if, in Box No. V, the name of the United States of America is accompanied by an indication "continuation" or "continuation-in-part": in such case, write "Continuation of Box No. V" and the name of each State involved (or OAPI), and after the name of each such State (or OAPI), the number of the parent title or parent application and the date of grant of the parent title or filing of the parent application;
- (vi) if, in Box No. VI, there are more than three earlier applications whose priority is claimed: in such case, write "Continuation of Box No. VI" and indicate for each additional earlier application the same type of information as required in Box No. VI;
- (vii) if, in Box No. VI, the earlier application is an ARIPO application: in such case, write "Continuation of Box No. VI", specify the number of the item corresponding to that earlier application and indicate at least one country party to the Paris Convention for the Protection of Industrial Property or one Member of the World Trade Organization for which that earlier application was filed.

2. If, with regard to the precautionary designation statement contained in Box No. V, the applicant wishes to exclude any State(s) from the scope of that statement: in such case, write "Designation(s) excluded from precautionary designation statement" and indicate the name or two-letter code of each State so excluded.

3. If the applicant claims, in respect of any designated Office, the benefits of provisions of the national law concerning non-prejudicial disclosures or exceptions to lack of novelty: in such case, write "Statement concerning non-prejudicial disclosures or exceptions to lack of novelty" and furnish that statement below.

Continuation of Box no. IV

RITTER, Stephen David)	All of: MATHYS & SQUIRE
GARRATT, Peter Douglas)	100 Gray's Inn Road
MOIR, Michael Christopher)	London
COZENS, Paul Dennis)	WC1X 8AL
SCHLICH, George William)	UNITED KINGDOM
COLMER, Stephen Gary)	
KAZI, Ilya)	
INGRAM, Brian Victor)	
SIMONS, Elisabeth Anne)	
BRADLEY, Josephine Mary)	
MacLEAN, Martin Robert)	

Box No. VI PRIORITY CLAIMS

 Further priority claims are indicated in the Supplemental Box.

Filing date of earlier application (day/month/year)	Number of earlier application	Where earlier application is:		
		national application: country	regional application: regional Office	international application: receiving Office
item (1) (22.02.99) 22 FEBRUARY 1999	9904028.9	GB		
item (2) (23.09.99) 23 SEPTEMBER 1999	9922561.7	GB		
item (3))			

The receiving Office is requested to prepare and transmit to the International Bureau a certified copy of the earlier application(s) (only if the earlier application was filed with the Office which for the purposes of the present international application is the receiving Office) identified above as item(s): (1) + (2)

* Where the earlier application is an ARIPO application, it is mandatory to indicate in the Supplemental Box at least one country party to the Paris Convention for the Protection of Industrial Property for which that earlier application was filed (Rule 4.10(b)(ii)). See Supplemental Box.

▲
Added
R/GS

Box No. VII INTERNATIONAL SEARCHING AUTHORITY

Choice of International Searching Authority (ISA)
(if two or more International Searching Authorities are competent to carry out the international search, indicate the Authority chosen; the two-letter code may be used):

ISA /

Request to use results of earlier search; reference to that search (if an earlier search has been carried out by or requested from the International Searching Authority):

Date (day/month/year) Number Country (or regional Office)

Box No. VIII CHECK LIST; LANGUAGE OF FILING

This international application contains the following number of sheets:

request	: 6
description (excluding sequence listing part)	: 22
claims	: 6
abstract	: 1
drawings	: 7
sequence listing part of description	: -
Total number of sheets	: 42

This international application is accompanied by the item(s) marked below:

1. fee calculation sheet
2. separate signed power of attorney
3. copy of general power of attorney, reference number, if any:
4. statement explaining lack of signature
5. priority document(s) identified in Box No. VI as item(s):
6. translation of international application into (language):
7. separate indications concerning deposited microorganism or other biological material
8. nucleotide and/or amino acid sequence listing in computer readable form
9. other (specify): 23/77 (x2)

Figure of the drawings which should accompany the abstract: 1

Language of filing of the international application: ENGLISH

Box No. IX SIGNATURE OF APPLICANT OR AGENT

Next to each signature, indicate the name of the person signing and the capacity in which the person signs (if such capacity is not obvious from reading the request).

SCHLICH, George William

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1. Date of actual receipt of the purported international application:	22 FEBRUARY 2000 22.2.2000	2. Drawings: <input checked="" type="checkbox"/> received: <input type="checkbox"/> not received:
3. Corrected date of actual receipt due to later but timely received papers or drawings completing the purported international application:		
4. Date of timely receipt of the required corrections under PCT Article 11(2):		
5. International Searching Authority (ISA / if two or more are competent):	6. <input type="checkbox"/> Transmittal of search copy delayed until search fee is paid.	

For International Bureau use only

Date of receipt of the record copy
by the International Bureau:

See Notes to the request form

NEISSERIAL VACCINE COMPOSITIONS AND METHODS

The present invention relates to vaccines and methods for preparing
5 vaccines that stimulate an immune response. In particular, the present invention relates to vaccines that provide broad spectrum protective immunity to microbial infection.

Infection by pathogenic organisms is one of the major causes of chronic and
10 acute disease. In particular, infection resulting from microbial sources - such as bacteria, viruses and protozoans - continue to claim millions of lives worldwide. With microbial species increasingly becoming resistant to conventional antibiotics, it would be desirable to provide alternative and preferably prophylactic means of protecting against and fighting microbial
15 infection.

Meningococcal meningitis is of particular importance as a worldwide health problem and in many countries the incidence of infection is increasing.
20 *Neisseria meningitidis* (the meningococcus) is the organism that causes the disease and is also responsible for meningococcal septicaemia, which is associated with rapid onset and high mortality, with around 22% of cases proving fatal.

At present, vaccines directed at providing protective immunity against
25 meningococcal disease provide only limited protection because of the many different strains of *N. meningitidis*. Vaccines based upon the serogroup antigens, the capsular polysaccharides, offer only short lived protection against infection and do not protect against many strains commonly found in North America and Europe. A further drawback of these vaccines is that
30 they provide low levels of protection for children under the age of 2 years, one of the most vulnerable groups that are commonly susceptible to infection. Newer conjugate vaccines now in use in the UK will address some

- 2 -

infection. Newer conjugate vaccines now in use in the UK will address some of these problems but will only be effective against the C serogroup of the meningococcus

5 Gold *et al.* (Journal of Infectious Diseases, volume 137, no. 2, February 1978, pages 112-121) have reported that carriage of *N. lactamica* may assist in the development of natural immunity to *N. meningitidis* by induction of cross-reactive antibodies. This conclusion was based on the observation of cross-reacting antibodies having complement-dependent bactericidal activity produced in response to *N. lactamica* infection. However, Cann and Rogers (J. Med. Microbiol., volume 30, 1989, pages 23-30) detected antibodies to common antigens of pathogenic and commensal neisseria species, but observed also that antibody to the same antigens was present in both bactericidal and non-bactericidal sera. Thus, it was not possible to 10 identify any cross-reactive bactericidal antibodies.
15

Live attenuated vaccines for meningococcal disease have been suggested by Tang *et al.* (Vaccine 17, 1999, pages 114-117) in which a live, attenuated strain of *N. meningitidis* could be delivered mucosally. Tang also commented on the use of commensal bacteria to protect against infection by pathogenic bacteria, concluding that the cross-reactive epitopes that induce protection against meningococcal infection have not been defined, and therefore that 20 use of genetically modified strains of *N. meningitidis* would be preferred.

25 It is desirable to provide a further vaccine that gives protective immunity to infection from *N. meningitidis*. It further is desirable to provide a vaccine that confers protective immunity to infants as well as adults and whose protection is long term. It may also be of advantage to provide a vaccine that protects against sub-clinical infection, i.e. where symptoms of meningococcal infection are not immediately apparent and the infected 30 individual may act as a carrier of the pathogen. It would further be of advantage to protect against all or a wide range of strains of *N. meningitidis*.

infection, notably gonorrhoea.

It is an object of the present invention to provide compositions containing immunostimulating components, and vaccines based thereon, that meet or
5 at least ameliorate the disadvantages in the art.

The present invention is based on the use of a commensal *Neisseria* in a vaccine against disease. Accordingly, a commensal species of *Neisseria* such as *N. lactamica* may be used as a live vaccine or a killed whole cell vaccine,
10 or in a vaccine containing fractions of *N. lactamica*. It has surprisingly been demonstrated that mice immunised according to the present invention with
N. lactamica killed whole cells and outer membrane preparations are protected from lethal intraperitoneal meningococcal challenge, and that
15 vaccines composed of a detergent extract of *N. lactamica* cells or fractions of this, separated by preparative electrophoresis, also protect mice from lethal meningococcal challenge. These results have been obtained using mice and the mouse model used is regarded as predictive of corresponding immunogenic and vaccinating effects in humans.

20 Accordingly, a first aspect of the present invention provides an immunogenic composition, comprising a commensal *Neisseria* or an immunogenic component, extract or derivative thereof and a pharmaceutically acceptable carrier.

25 The composition of the invention is particularly suited to vaccination against infection of an animal. The term "infection" as used herein is intended to include the proliferation of a pathogenic organism within and/or on the tissues of a host organism. Such pathogenic organisms typically include bacteria, viruses, fungi and protozoans, although growth of any microbe within and/or on the tissues of an organism are considered to fall within the
30 term "infection".

- 4 -

Commensal micro-organisms are those that can colonize a host organism without signs of disease. A number of different commensal *Neisseria* are suitable for use in the invention, and these commensal *Neisseria* may be selected from the group consisting of *N. lactamica*, *N. cinerea*, *N. elongata*,
5 *N. flavescens*, *N. polysaccharea*, *N. sicca* and *N. subflava*. Different species of these commensal organisms are known to colonise the buccal or nasal areas or other mucosal surfaces and hence each species may be administered according to the known area of the body it normally colonises.
10 Hence also, use of a composition of the invention may result in stimulation of production of protective antibodies *de novo* or if the individual has already been colonised to a certain extent may result in an enhancement of naturally-existing antibodies.

15 The "extract" or "component" is an extract or component that is immunogenic such that antibodies raised against the extract or component of a commensal *Neisseria* cross react with a pathogenic *Neisseria*, in particular *N. meningitidis*.

20 The term "derivative" is used to describe types and strains of commensal *Neisseria* that are modified or attenuated in some way so as to differ from the wild type species; for example, a vaccine composition comprising a recombinant commensal *Neisseria* that exhibits resistance to certain types of antibiotic compounds, which might advantageously be utilised in combination with such antibiotics in the treatment of infection.

25 It is an advantage of the invention that vaccination against neisserial diseases may thus be achieved using a non-pathogenic species of *Neisseria*, rendering the vaccination a safer procedure. Furthermore, the protection conferred surprisingly may not be restricted to a specific serotype, subtype
30 or serogroup of the meningococcus but is of general protective efficacy.

A further advantage of the invention is that the commensal *Neisseria* that are

- 5 -

the subject of the invention can not revert to virulent types. It is known in the vaccination field to use live, attenuated pathogens and this use carries the risk that the attenuated organism may revert to virulence. This risk is avoided by the present invention. Furthermore, *N. meningitidis* possesses many virulence factors the precise roles of which in pathogenesis are unknown and may possess hitherto unrecognised virulence factors. Therefore, an additional advantage of the invention is that a composition of the invention can be used with confidence in its level of safety.

10 The method of the invention is of application to vaccination against various infections, preferably but not only neisserial infections. In a specific embodiment of the invention, protection against meningococcal disease has been demonstrated. The invention is also of application to vaccination generally against neisserial infection, including gonorrhoeal infection, and
15 also to infection from other pathogenic microbial organisms. The invention further provides for vaccination that is aimed at either stimulating or desensitizing the immune system.

20 The composition can specifically comprise killed commensal *Neisseria*, which may for example be obtained by heat or by suspending commensal *Neisseria* in a mixture of bactericidal agents such as thiomersal and formaldehyde.

25 The composition may also comprise live commensal *Neisseria*. As mentioned, it is optional but not usually required to use attenuated commensal *Neisseria* as these organisms are avirulent.

30 In an embodiment of the invention, an immunogenic component or extract of a commensal *Neisseria* is selected from an outer membrane vesicle preparation, an outer membrane preparation, lipooligosaccharide and a protein fraction.

The outer membrane preparation and protein fraction of *N. lactamica*, for

example, can be obtained from *N.lactamica* cultured in the presence or absence of iron. The protein fraction of *N.lactamica* is conveniently obtained by suspending *N.lactamica* cells or membranes in the presence of detergent and incubating the suspension so as to extract proteins from the *N.lactamica*.

5

Alternatively, a number of other techniques are known for extraction of outer membrane components - such as protein fractions, lipooligosaccharides and lipopolysaccharides - from cell preparations and are suitable to obtain the commensal *Neisseria* immunogenic components or extracts of the invention. Examples of conventional techniques for this purpose include the use of variation in salt concentration, chaotropic agents, variation in pH (high or low), enzymic digestion and mechanical disruption.

10

15

A number of different fractions are suitable for use in vaccinating against meningococcal disease. Particularly suitable fractions are those of molecular weight less than 50kDa, of molecular weight more than 40kDa and less than 70kDa, and of molecular weight more than 60kDa.

20

In more specific embodiments of the invention there is provided a composition for eliciting an immune response and suitable for use in vaccinating an individual against neisserial infection, more specifically meningococcal disease, comprising an antigenic component or antigenic components having the properties:-

25

- (a) molecular weight 50kDa or lower;
- (b) obtainable from *N. lactamica*; and
- (c) antibodies to the component(s) obtained from *N. lactamica* cross-react with *N. meningitidis*.

30

In use of a composition containing such a component, extracted using detergent, all mice treated with this component survived a challenged dose of 2×10^7 CFU *N. meningitidis* and three out of five mice survived a higher

challenge dose of 6×10^8 CFU.

Another specific embodiment of the invention lies in a composition for eliciting an immune response and suitable for use in vaccinating an individual against neisserial infection, more specifically meningococcal disease, comprising an antigenic component or antigenic components having the properties:-

- (a) molecular weight at least 40kDa and up to 70kDa;
(b) obtainable from *N. lactamica*; and
10 (c) antibodies to the component(s) obtained from *N. lactamica* cross-react with *N. meningitidis*.

In use of such a component of the invention, obtained using a detergent extract of *N. lactamica*, four out of five mice treated with the component survived a challenge dose of 2×10^7 CFU *N. meningitidis* and mice receiving a higher challenge dose of 6×10^8 CFU survived longer than a control group.

A still further embodiment of the invention lies in a composition for eliciting an immune response and suitable for use in vaccinating an individual against neisserial infection, more specifically meningococcal disease, comprising an antigenic component or antigenic components having the properties:-

- (a) molecular weight at least 60kDa;
(b) obtainable from *N. lactamica*; and
20 (c) antibodies to the component(s) obtained from *N. lactamica* cross-react with *N. meningitidis*.

In use of such a component, obtained using a detergent extract, one out of five mice survived a challenge dose of 2×10^7 CFU *N. meningitidis* and, whilst all mice succumbed to a higher challenged dose of 6×10^8 CFU, their survival time was longer than a control group which did not receive the component.

In an example of the invention in use, described in more detail below, proteins in the size ranges of 25-35 kDa and 35-43 kDa, extracted from a commensal *Neisseria*, conferred a significant level of immune protection when administered to mice as a vaccine composition.

5

By way of example of a method of extracting an antigenic component of the invention, an extraction method comprises:-

- 10 (i) suspending *N.lactamica*, cells in an aqueous solution of detergent;
- (ii) incubating the suspension so as to extract the antigenic component from the *N.lactamica*;
- 15 (iii) centrifuging the suspension to separate the suspension into a supernatant and a pellet; and
- (iv) fractionating the antigenic component from the supernatant.

20 This specific method can be modified according to the extraction protocol selected by the user, for example by using high salt concentration in the initial step (i). In further embodiments of the invention the antigenic component is obtained using recombinant technology by expression of a *N.lactamica* sequence in a suitable host such as *E. coli*.

25

In a second aspect of the invention there is provided a composition for vaccination against neisserial infection comprising a commensal *Neisseria* or an immunogenic component, extract or derivative thereof and a pharmaceutically acceptable carrier, wherein the commensal *Neisseria* comprises and expresses a gene from a pathogenic *Neisseria*.

30 This aspect of the invention offers the benefit of use of a commensal

organism to deliver and/or present to the recipient an antigen from a pathogenic *Neisseria*. The gene optionally encodes a surface antigen or a protein that is secreted, and may code for an antigen from, for example, *N. meningitidis* or *N. gonorrhoeae*. The commensal *Neisseria* can be live or
5 killed.

In an embodiment of the second aspect of the invention there is provided a composition for vaccination against meningococcal disease comprising a commensal *Neisseria* and a pharmaceutically acceptable carrier, wherein the
10 commensal *Neisseria* comprises and expresses a *N. meningitidis* gene.

The *N. meningitidis* gene may encode for example a transferrin binding protein, a superoxide dismutase (SOD) for example a Cu,Zn SOD, neisserial surface protein A ("NspA"), a porin or another outer membrane protein.
15 Gene sequences for the majority of these antigens are known in the literature. Kroll *et al.* in Microbiology 141 (Pt 9), 2271-2279 (1995) describe the sequence of Cu,Zn-SOD. Martin *et al.* in J Exp Med, 1997, April 7th, 185(7), pp1173-1183 describe the sequence of NspA from *N. meningitidis*.

20 The invention also provides a pharmaceutical composition comprising a composition according to the first or second aspect of the invention plus a pharmaceutically acceptable carrier.

In a third aspect, the invention provides a method of vaccination against
25 neisserial infection, comprising administering an effective amount of a composition according to the first and second aspects of the invention.

30 In use of an embodiment of the invention described in an example below, there is provided a method of vaccination against meningococcal disease, comprising administering an effective amount of a composition according to the first and second aspects of the invention.

- 10 -

In a fourth aspect of the invention there is provided a strain of a commensal *Neisseria*, such as *N. lactamica*, genetically modified so as to express a gene from a pathogenic *Neisseria*. The *N.meningitidis* gene may for example code for a protein selected from a transferrin binding protein, a SOD for example a Cu,Zn-SOD, NspA, a porin or another outer membrane protein.

5

The invention further provides, in a fifth aspect a method of extracting a protein for incorporation in a composition suitable for vaccinating against meningococcal disease, comprising:-

10

- (i) suspending commensal *Neisseria*, for example *N.lactamica*, cells in the presence of detergent; and

- (ii) incubating the suspension so as to extract a protein fraction from the cells.

15

The protein fraction can suitably be of molecular weight 50kDa or lower, at least 40kDa and up to 90kDa or at least 80kDa.

20

The composition may be combined with a pharmaceutically acceptable carrier - for example the adjuvant alum although any carrier suitable for oral, intravenous, subcutaneous, intraperitoneal intramuscular, intradermal or any other route of administration is suitable - to produce a pharmaceutical composition for treatment of meningococcal disease. Commensal *Neisseria* that are buccal colonizers can be administered in a mouthwash and nasal colonizers in a nasal spray.

25

Transferrin binding proteins are known to be located on the outer membranes of a number of Gram negative bacteria such as *N. meningitidis*.

30

Formulations of the composition of the present invention with conventional carriers or adjuvants and optionally further supplemented by one or more antigens from *Neisseria* species, optionally recombinantly produced, for

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example, Cu-Zn SOD, the 22kD NspA, porins, gonorrhoeal antigens or transferrin binding proteins provide a composition for treatment of infection by these bacteria.

- 5 In the present invention, the term "transferrin binding protein" or "Tbp" refers to a protein which either alone binds to transferrin or can be part of a complex of proteins that binds transferrin. The term also embraces fragments, variants and derivatives of such a protein provided that antibodies raised against the fragment, variant or derivative bind the protein.
- 10 Thus, TbpA and TbpB either dissociated or associated into a complex are considered to be Tbp. Moreover, mutants, fusion proteins or fragments of either TbpA or B or other derivatives of the TbpA+B complex with a common antigenic identity are also considered to be represented by the term Tbp in the present invention.
- 15 A live vaccine according to the present invention may be administered parenterally or to the mucosa for example via intranasal or oral inoculation. A killed bacteria or subunit vaccine may also be given by this route, or formulated for oral delivery. A subunit vaccine is conveniently administered via the parenteral route. Different commensal *Neisseria* and different strains of *N. lactamica* from those tested in specific embodiments of the invention exist, and the invention is of application also to those other strains.
- 20 A sixth aspect of the invention provides a composition comprising an antibody, wherein the antibody binds to a commensal *Neisseria* of the first or second aspects of the invention or an immunogenic component or extract thereof. In use, the antibody can be formulated into a pharmaceutical composition for treatment of neisserial infection, such as meningococcal disease or infection caused by other *Neisseria*.
- 25 An antibody according to this aspect of the invention can be obtained following standard techniques, for example by inoculating an animal with the

commensal *Neisseria* or an immunogenic component or extract thereof and thereafter isolating antibodies that bind to the commensal *Neisseria* or the immunogenic component or extract thereof.

- 5 A further aspect of the invention provides for a composition comprising a commensal *Neisseria*, or an immunogenic component, extract or derivative thereof, wherein said *Neisseria* comprises a heterologous gene product.

10 Heterologous gene products of the invention typically include peptides, proteins and antisense sequences that are coded for by a gene sequence that is not native to the commensal *Neisseria*. Typical heterologous gene products of the invention include, for example, bacterial proteins, viral proteins or surface peptides, antigens and antibodies and fragments thereof. The heterologous gene product of the invention may also be any antigen found in a pathogenic organism.

15 In an embodiment of the invention, the composition comprises a commensal *Neisseria* into which has been transformed an expression vector containing a gene sequence encoding a heterologous gene product. Specific proteins suitable for use in the invention typically include:-

20 Viral proteins - such as hepatitis B virus surface antigen; rabies virus glycoprotein G; herpes simplex virus glycoprotein D; Epstein-Barr virus glycoprotein; influenza virus glycoprotein; vesicular stomatitis virus nucleoprotein; human respiratory syncytial virus glycoprotein G; human immunodeficiency virus (HIV) envelope; rotavirus subunits; measles virus subunits; and vaccinia virus subunits.

25 Bacterial proteins - such as *Bordetella pertussis* fimbrial subunits; *Bordetella pertussis* surface proteins; *Bacillus anthracis* subunits; *Escherichia coli* subunits; and *Yersinia pestis* subunits.

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Protozoan proteins - such as *Plasmodium falciparum* proteins; trypanosome proteins; and *Cryptosporidium* proteins.

In a further embodiment the composition of the invention suitably provides
5 for a commensal *Neisseria* that expresses a heterologous gene product which is immunostimulatory for treatment of non-infectious disease, for example allergy and cancer. In an example of the invention in use a commensal *Neisseria* that expresses peanut antigens is used to desensitize a patient with acute peanut allergy.

10 In a further example of the invention in use, described in more detail below, the expression vector pJSK422 is used to express green fluorescent protein, under the control of the groES/EL promoter, in the commensal *N. cinerea*.

15 The invention further provides for a commensal *Neisseria* that is transformed with an expression vector that comprises a signal sequence that directs the heterologous gene product to the outer membrane of the neisserial cell. Other signal sequences are also suitable for use in the invention, such as secretion signals or cellular subcompartment localisation signals e.g. 20 periplasmic localisation signals.

Further aspects of the invention provide methods for preparing compositions. Such methods are suitable for preparing vaccine compositions that elicit protective immunity to microbial infection when administered to an animal.

25 An example of the invention in use, described in more detail below, provides for a method of preparing a composition comprising the steps of:

- a) inserting a gene coding for a heterologous gene product into an expression vector;
- 30 b) transforming said expression vector into a commensal

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Neisseria so that said heterologous gene product is expressed in said *Neisseria*; and

- 5 c) combining the *Neisseria* of (b) with a pharmaceutically acceptable carrier.

A further example of the invention, provides for a method of preparing a composition comprising the steps of:

- 10 a) inserting a gene coding for a heterologous gene product into an expression vector;
- 15 b) transforming said expression vector into a commensal *Neisseria* so that said heterologous gene product is expressed in said *Neisseria*;
- 20 c) obtaining an immunogenic component or extract from the *Neisseria* of (b); and
- 25 d) combining the immunogenic component or extract of (c) with a pharmaceutically acceptable carrier.

In yet a further example of the invention in use is provided a method of preparing a composition comprising the steps of:

- 25 a) obtaining an immunogenic component or extract from a commensal *Neisseria*; and
- 30 b) combining the immunogenic component or extract of (a) with a heterologous gene product and a pharmaceutically acceptable carrier.

- 15 -

Thus, the invention provides for (a) methods and compositions in which an extract is taken from a commensal *Neisseria* that expresses a heterologous gene product, and (b) methods and compositions where an extract is obtained from a commensal *Neisseria* and the heterologous gene product expressed elsewhere (in another organism) is combined with this latter extract.

Further aspects of the invention provide for use of a commensal *Neisseria* in the manufacture of a medicament for treatment of neisserial infection, and for use of a commensal *Neisseria*, or an immunogenic component, extract or derivative thereof, wherein said *Neisseria* comprises a heterologous gene product, in the manufacture of a medicament for the treatment of infection or for immunostimulation in an animal.

Specific embodiments of the invention are discussed in more detail by means of the Examples described below. The results referred to in the Examples are illustrated by the accompanying drawings, in which:

Fig. 1 shows protection of mice against intraperitoneal ("IP") infection with *N.meningitidis* strain K454 by use of *N.lactamica* whole cells and outer membrane fractions;

Fig. 2A shows protection of mice against IP infection with *N.meningitidis* strain K454 by use of detergent and high, medium and low molecular weight extracts of *N.lactamica* cells - upper panel = challenge by 2×10^7 CFU, lower panel = challenge by 6×10^8 CFU;

Fig. 2B shows the components of the high, medium and low molecular weight fractions of fig.2A;

Fig. 3 shows an immunoblot illustrating cross-reaction of antibodies in sera from meningococcal disease patients with proteins from *N.lactamica*

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strain Y92-1009;

Fig. 4 shows a photograph of a gel on which subfractions of low molecular weight outer membrane protein extract have been run; and

5

Fig. 5 shows protection of mice against IP infection with *N.meningitidis* strain K454 when immunised with low molecular weight subfractions - upper panel=challenge by 5×10^6 CFU, lower panel=challenge by 1×10^8 CFU.

10

Fig. 6 shows a histogram comparing the fluorescence of *N. cinerea* NRL 32165 containing pJSK411 (promoterless GFP) to pJSK422 (pJSK411 with groEL/ES promoter).

15

Example 1

Preparation of vaccine containing killed whole cells

20 *Neisseria lactamica* strain Y92-1009 was grown in Mueller Hinton broth (MHB) containing $5\mu\text{gml}^{-1}$ ethylenediamine-di(o-hydroxyphenylacetic acid) (EDDHA), incubated at 37°C with shaking (140rpm) for approximately 6h.

25

Bacteria were then harvested by centrifugation and resuspended in phosphate buffered saline (PBS) containing 1% (v/v) formaldehyde and 0.1% (w/v) thiomersal, and left to stand overnight at 2-8°C. Killed cells were then resuspended in PBS to an OD_{650} of 1.0 (equivalent to $2 \times 10^9 \text{ CFUml}^{-1}$) and alhydrogel added to 25% (V/V), yielding a product suitable for subcutaneous administration.

30

This method is suitable also for *N. cinerea*, *N. elongata*, *N. flavescens*, *N. polysaccharea*, *N. sicca* and *N. subflava*.

Example 2**Preparation of vaccine containing *N. lactamica* outer membrane (OM) preparations**

5 *N. lactamica* strain Y92-1009 was grown in MHB with and without the addition of $5\mu\text{gml}^{-1}$ EDDHA overnight at 37°C with shaking. Iron limited (with EDDHA) and iron replete cells were then treated separately. Bacteria from 1.5 litres were harvested by centrifugation and resuspended in 20ml
10 200mM Lithium acetate, 5mM EDTA, pH 6.0 and incubated for 3h at 37°C with shaking. Bacteria were then passed 7 times through a 21 gauge needle and pelleted at 8000g for 10min.

15 The supernatant was recovered and membranes pelleted by centrifugation at 100,000g for 1h at 4°C. The membranes were then resuspended in 10mM HEPES, pH 7.4, containing 0.1% (v/v) 10mM PMSF, yielding OM-
20 containing vaccinating preparations. The protein content of the OM vaccine preparations was determined using the bicinchoninic acid assay (Sigma, UK). OMs were diluted in sterile deionized water to give a protein concentration of $100\mu\text{gml}^{-1}$. This was then mixed with an equal volume of Freund's adjuvant, to give a final protein concentration of $50\mu\text{gml}^{-1}$, and emulsified thoroughly. Freund's complete adjuvant was used for the primary dose, and Freund's incomplete for subsequent boosts.

25 Example 3**Preparation of vaccine containing lipooligosaccharide (LOS)**

30 Purification of LOS was carried out from *N. lactamica* strain Y92-1009 using the method of Gu, X-X and Tsai, C.M. (1991) Anal Biochem. 196; 311-318. Vaccine was prepared using Freund's adjuvant as above with LOS at a final concentration of $10\mu\text{gml}^{-1}$.

Example 4**Vaccination and challenge schedule**

5 Groups of 5 mice were vaccinated with each preparation as follows:-

Prime:-	Day 0
First boost:-	Day 21
Second boost:-	Day 28

10 Mice vaccinated with killed cells of Example 1 received 0.5ml subcutaneously, equivalent to 1×10^9 CFU. Mice vaccinated with OM of Example 2 and LOS of Example 3 received 0.2ml subcutaneously; equivalent to 10 μ g of protein and 2 μ g of LOS.

15 On day 35, mice were challenged by intraperitoneal injection with approximately 10^8 CFU *N. meningitidis* K454 made up in MHB containing transferrin at a final concentration of 20mg/ml. The mice were then examined and the number of survivors noted and the results are shown in fig.1. After 4 days all 5 mice survived in the groups vaccinated with whole cells and OMPs (without iron) and 3 survived in the group vaccinated with OMPs (with iron). After 5 days all members of the control group and of the group vaccinated with LOS (marked LPS on the figure) had died.

Example 5

25

Preparation of vaccine comprising *N. lactamica* fractions

30 Brain heart infusion agar plates were inoculated with 50 μ l of *N. lactamica* strain Y92-1009 and incubated overnight at 37°C, with 5% CO₂. This was used to inoculate a 100ml MHB starter culture which was incubated with shaking at 37°C for 6 h. Starter culture (15ml) was added to each of 6x500ml volumes of MHB. These were then incubated with shaking

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overnight at 37°C and the conditions were made iron-limited by the addition of 5ug/ml¹ EDDHA. The cells were harvested by centrifugation and the supernatant discarded. The cells were washed with 100ml PBS and then pelleted by centrifugation. Cell pellets were resuspended in PBS + 0.3% (v/v) Elugent (Calbiochem, 2ml per g wet weight) and incubated with shaking at 37°C for 20 min. The cells were then removed by centrifugation and the pellet discarded. EDTA and N-lauroyl sarcosine were then added to the supernatant to 10mM and 0.5% (w/v) respectively.

The BioRad (Registered Trade Mark) Prep Cell, model 491 was then used to separate the proteins contained in the detergent extract. A 4cm, 7% acrylamide native resolving gel was cast with a 2 cm stacking gel. 12mg of protein in native sample buffer was electrophoresed using running buffer containing 0.1% (w/v) SDS, 0.025M Tris and 0.192M glycine at 40mA and 400V until the dye front reached the bottom of the gel. 3ml fractions of the eluted proteins were then collected. Once the fractions were collected they were pooled into groups consisting of proteins of molecular weight approximately less than 40kDa, between 40 and 67kDa and more than 67kDa. The pooled proteins were concentrated by ammonium sulphate precipitation and dialysed against PBS. These were diluted in PBS to a protein concentration of 100ug/ml and Freund's complete adjuvant was added at a ratio of 1:1(v/v) or Freund's incomplete adjuvant for booster doses.

Example 6

Vaccination and challenge schedule

Groups of 5 mice were vaccinated with each preparation as follows:-

Prime:- Day 0
First boost:- Day 21
Second boost:- Day 28

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Mice were vaccinated with no vaccine (i.e control group), Elugent ("Registered Trade Mark") extract or high, medium or low molecular weight fraction. The mice receiving the protein fraction groups received 0.2ml subcutaneously; equivalent to 10 μ g of protein.

5

On day 35 mice were challenged by intraperitoneal injection with either approximately 2×10^7 or 6×10^8 CFU *N. meningitidis* K454 made up in MHB containing transferrin at a final concentration of 20mg/ml. The mice were then examined over four days and the number of survivors noted, and the results are shown in fig.2A - upper panel 2×10^7 challenge and lower panel 6×10^8 challenge. The components of the high, medium and low molecular weight fractions are shown in fig.2B. after being run an SDS-PAGE gel.

10

Example 7

15

Samples of human sera following meningococcal disease were investigated and these showed that antibodies were produced which react with a range of *N. lactamica* proteins. The results of the immunoblot are shown in fig.3.

20

Example 8

25

Due to the level of protection offered by the low molecular weight pool in Example 6 (see Fig. 2A), further separation of these proteins was carried out, according to the method of Example 5, to further characterise components responsible for protection. Proteins were pooled into three groups consisting of <25 kDa (g1), 25-35 kDa (g2) and 35-43 kDa (g3)(shown in Fig. 4). Determination of the levels of lipopolysaccharide (LPS) revealed high levels of LPS in fraction g1 [26 580 endotoxin units per ml (EUml $^{-1}$)], and considerably lower levels in the remaining fractions (9149 EUml $^{-1}$ in g2 and 9348 EUml $^{-1}$ in g3).

30

As in previous examples, groups of five mice were immunised, using a three

- 21 -

dose schedule with one of the three groups of proteins described above, proteins >43 kDa and detergent extract of killed whole *N. lactamica* cells and killed whole *N. lactamica*. Animals were challenged with *N. meningitidis* serogroup B, strain K454, at a dose of 5×10^6 or 1×10^8 CFU, together with unimmunised controls. The number of survivors on each day post challenge 5 is shown in Fig. 5.

All mice, apart from the control group and one mouse in group g3, survived the lower challenge dose; however, at the higher challenge dose the g2 and 10 g3 protein groups (25-35 kDa and 35-43 kDa respectively) offered best protection.

Example 9

15 **Commensal *Neisseria* as a vehicle for recombinant protein expression**

The gene encoding the measles virus nucleocapsid protein was cloned into 20 the pMGC18.1 shuttle vector (Webb *et al.*, 1998, poster at the 11th International Pathogenic Neisseria Conference, Nice) and transformed into *E.coli* DH5alpha. Expression of the measles virus nucleocapsid protein was confirmed by western blotting probed with specific antiserum. This construct was then used to transform *N. lactamica* by conjugation. Expression of the 25 measles virus nucleocapsid protein was placed under the control of the neisserial *frpC* promoter and expression at high levels was seen when the bacteria were grown under iron-limited growth conditions.

Example 10

30 **Expression of GFP in the commensal *N. cinerea***

The green fluorescent protein (GFP) gene of *Aequorea victoria* was inserted

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into the pJSK422 plasmid using standard cloning techniques. The GFP was under the control of the groES/EL promoter. As a negative control the GFP gene was also inserted into the pJSK411 plasmid which lacks the groES/EL promoter of the pJSK422 plasmid.

5

N. cinerea was transformed via conjugation (see Example 9) either with the pJSK422 or pJSK411 (negative control) GFP containing plasmids. The transformed cells were cultured under appropriate conditions. Fluorescence of the pJSK422 transformed cultures of *N. cinerea* were compared to that of the pJSK411 transformed cultures. The results of the comparison are shown in Fig. 6. The histogram shows intensity of GFP fluorescence on the X axis and the number of cells fluorescing on the Y axis. It is clear that the level of fluorescence is higher in the *N. cinerea* transformed with pJSK422 than those transformed with pJSK411, indicated by the peak shift to the right. This, demonstrates heterologous expression of the GFP gene in the commensal *N. cinerea*.

10

15

The invention thus provides immunogenic compositions and vaccines for use in protecting against disease.

CLAIMS

1. A vaccine composition comprising a commensal *Neisseria* or an immunogenic component, extract or derivative thereof.
2. A composition according to Claim 1 comprising killed *Neisseria*.
3. A composition according to Claim 2 wherein killed *N.lactamica* are obtained by suspending *N.lactamica* in a mixture of bactericidal agents such as thiomersal and formaldehyde.
4. A composition according to Claim 1 comprising live *N. lactamica*.
5. A composition according to Claim 1 wherein the immunogenic component or extract of *Neisseria* is selected from an outer membrane preparation, a lipooligosaccharide and a protein fraction.
6. A composition according to Claim 5 wherein the *Neisseria* is *N. lactamica*.
7. A composition according to Claim 5 wherein the immunogenic component or extract comprises a protein fraction of molecular weight less than 40kDa.
8. A composition according to Claim 5 wherein the immunogenic component or extract comprises a protein fraction of molecular weight more than 40kDa and less than 67kDa.
9. A composition according to Claim 5 wherein the immunogenic component or extract comprises a protein fraction of molecular weight more than 67kDa.
10. A composition according to any of Claims 7 to 9 wherein a protein

fraction of *N.lactamica* is obtained by suspending *N.lactamica* cells in the presence of detergent and incubating the suspension so as to extract proteins from the *N.lactamica*.

11. A composition for eliciting an immune response and suitable for use in vaccinating an individual against neisserial infection, comprising an antigenic component having the properties:-
 - (a) it is of molecular weight 50kDa or lower;
 - (b) it is obtainable from a commensal *Neisseria*; and
 - (c) antibodies to the component obtained from the commensal *Neisseria* cross-react with *N. meningitidis*.
12. A composition for eliciting an immune response and suitable for use in vaccinating an individual against neisserial infection, comprising an antigenic component having the properties:-
 - (a) it is of molecular weight at least 40kDa and up to 90kDa;
 - (b) it is obtainable from a commensal *Neisseria*; and
 - (c) antibodies to the component obtained from the commensal *Neisseria* cross-react with *N. meningitidis*.
13. A composition for eliciting an immune response and suitable for use in vaccinating an individual against neisserial infection, comprising an antigenic component having the properties:-
 - (a) it is of molecular weight at least 70kDa;
 - (b) it is obtainable from a commensal *Neisseria*; and
 - (c) antibodies to the component obtained from the commensal *Neisseria* cross-react with *N. meningitidis*.
14. A composition according to any of Claims 11 to 13 further comprising one or more of a transferrin binding protein, a Cu,Zn-SOD, a porin and NspA.
15. A composition for vaccination against neisserial infection comprising a

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commensal *Neisseria* and a pharmaceutically acceptable carrier, wherein the commensal *Neisseria* expresses a gene from a pathogenic *Neisseria*.

16. A composition according to Claim 15 wherein the commensal *Neisseria* expresses a gene which encodes a protein from *N. meningitidis* selected from the group consisting of transferrin binding protein; a Cu,Zn-SOD; an NspA; a porin; and an outer membrane protein.
17. A pharmaceutical composition comprising a composition according to any of Claims 1 to 14 plus a pharmaceutically acceptable carrier.
18. A method of vaccination against microbial infection, comprising administering an effective amount of a composition according to any of Claims 1 to 17.
19. A method of vaccination against neisserial infection, comprising administering an effective amount of a composition according to any of Claims 1 to 17.
20. A commensal *Neisseria*, which expresses a gene from a pathogenic *Neisseria*.
21. A commensal *Neisseria* according to Claim 20, which expresses a *N.meningitidis* gene which codes for a protein selected from the group consisting of: a transferrin binding protein, a porin, NspA, an outer membrane protein, and a Cu,Zn-SOD.
22. A method of extracting a protein for incorporation in a composition suitable for vaccinating against meningococcal disease, comprising:-
 - (i) suspending commensal *Neisseria* cells in the presence of detergent; and

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- (ii) incubating the suspension so as to extract a protein fraction from the cells.
23. A method according to Claim 22, wherein the protein fraction is of molecular weight 50kDa or lower.
24. A method according to Claim 22, wherein the protein fraction is of molecular weight at least 40kDa and up to 90kDa.
25. A method according to Claim 22, wherein the protein fraction is of molecular weight at least 80kDa.
26. A method according to any of Claims 22 to 25 further comprising mixing the protein fraction with one or more of a transferrin binding protein, a Cu,Zn-SOD, a porin, an NspA and a meningococcal protein.
27. A pharmaceutical composition comprising an antibody that binds to a commensal *Neisseria* or an immunogenic component or extract thereof and a pharmaceutically acceptable carrier.
28. A composition for vaccinating against meningococcal disease substantially as hereinbefore described.
29. A composition comprising a commensal *Neisseria*, or an immunogenic component, extract or derivative thereof, wherein said *Neisseria* comprises a heterologous gene product.
30. A composition according to Claim 29 wherein the heterologous gene product is a product of a gene that is expressed in said commensal *Neisseria*.
31. A composition according to Claim 29 wherein the heterologous gene

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product is physically combined with said commensal *Neisseria*.

32. A method of preparing a composition comprising:

- a) inserting a gene coding for a heterologous gene product into an expression vector;
- b) transforming said expression vector into a commensal *Neisseria* so that said heterologous gene product is expressed in said *Neisseria*; and
- c) combining the *Neisseria* of (b) with a pharmaceutically acceptable carrier.

33. A method of preparing a composition comprising the steps of:

- a) inserting a gene coding for a heterologous gene product into an expression vector;
- b) transforming said expression vector into a commensal *Neisseria* so that said heterologous gene product is expressed in said *Neisseria*;
- c) obtaining an immunogenic component or extract from the *Neisseria* of (b); and
- d) combining the immunogenic component or extract of (c) with a pharmaceutically acceptable carrier.

34. A method of preparing a composition comprising the steps of:

- a) obtaining an immunogenic component or extract of a

commensal *Neisseria*; and

- b) combining the immunogenic component or extract of (a) with a heterologous gene product and a pharmaceutically acceptable carrier.
35. A method according to any of claims 29-34, wherein said commensal *Neisseria* is selected from the group consisting of *N. lactamica*; *N. cinerea*; *N. elongata*; *N. flavescens*; *N. polysaccharea*; *N. sicca*; and *N. subflava*.
35. Use of a commensal *Neisseria* in the manufacture of a medicament for treatment of neisserial infection.
36. Use of a commensal *Neisseria*, or an immunogenic component, extract or derivative thereof, wherein said *Neisseria* comprises a heterologous gene product, in the manufacture of a medicament for the treatment of infection in an animal.
37. Use of a commensal *Neisseria*, or an immunogenic component, extract or derivative thereof, wherein said *Neisseria* comprises a heterologous gene product, in the manufacture of a medicament for immunostimulation in an animal.

NEISSERIAL VACCINE COMPOSITIONS AND METHODS**ABSTRACT**

Methods and compositions for the treatment of microbial infection, and in particular meningococcal disease, comprise a commensal *Neisseria* or an extract of a commensal *Neisseria*. Further methods and compositions comprise commensal *Neisseria* which express genes from virulent strains of *Neisseria* and/or heterologous gene products from non-neisserial sources. Such compositions are used in vaccine preparations for the treatment of microbial infection.

[Fig. 1]

Fig. 1. Protection of mice against IP infection with *N. meningitidis* strain K454 by vaccination with *N. lactamica* whole cells and outer membrane proteins

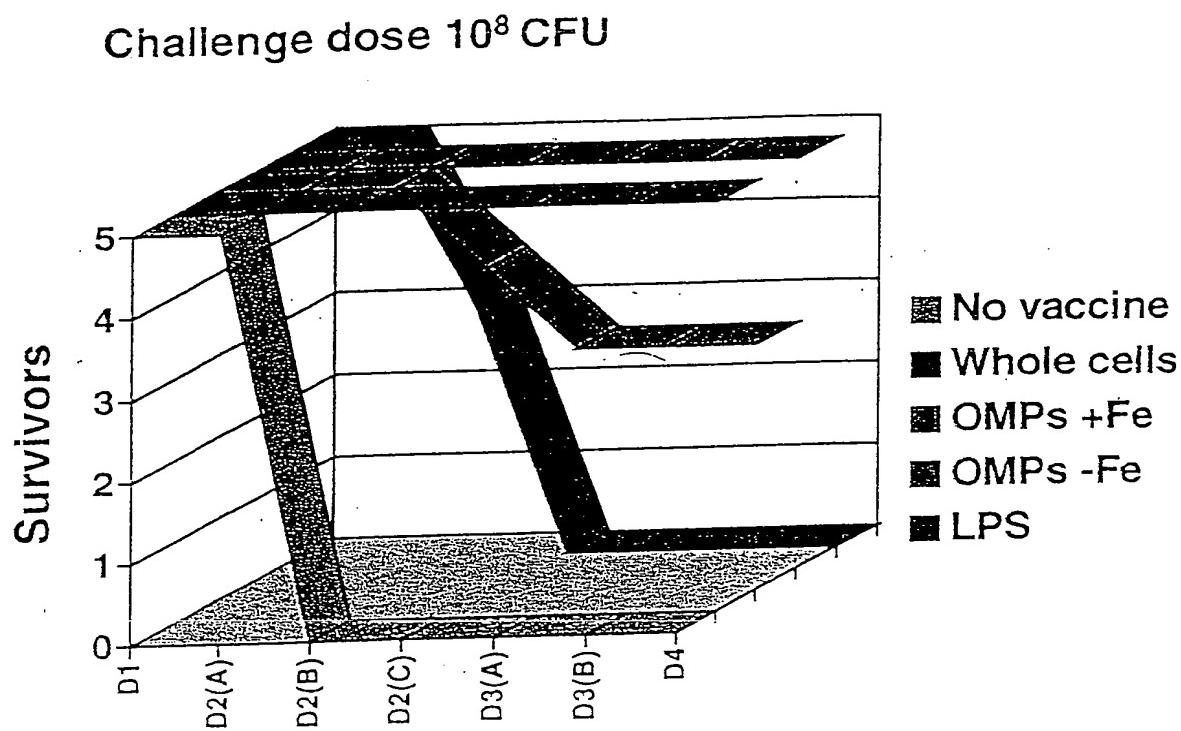


Fig. 2A. Protection of mice against IP infection with *N. meningitidis* strain K454 by vaccination *N. lactamica* extracts

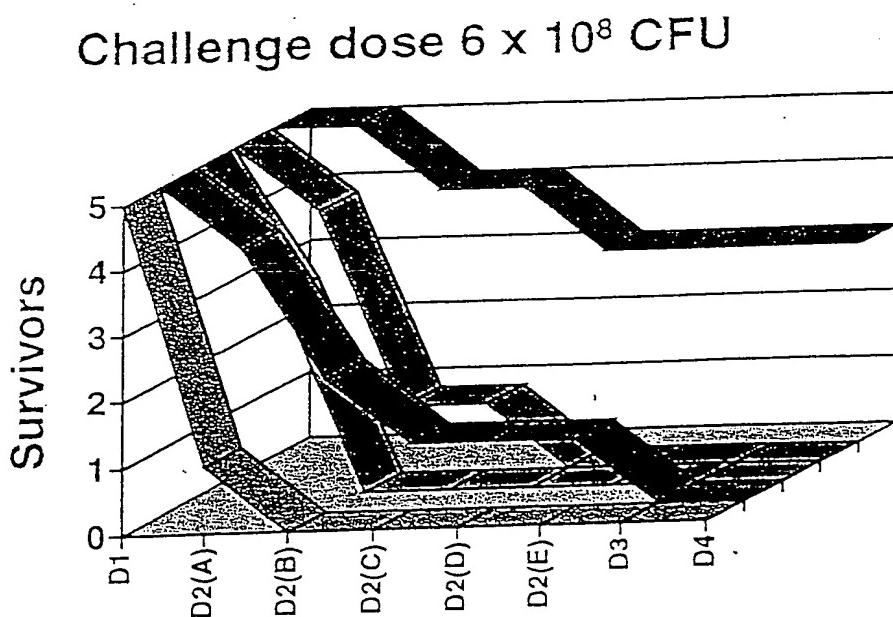
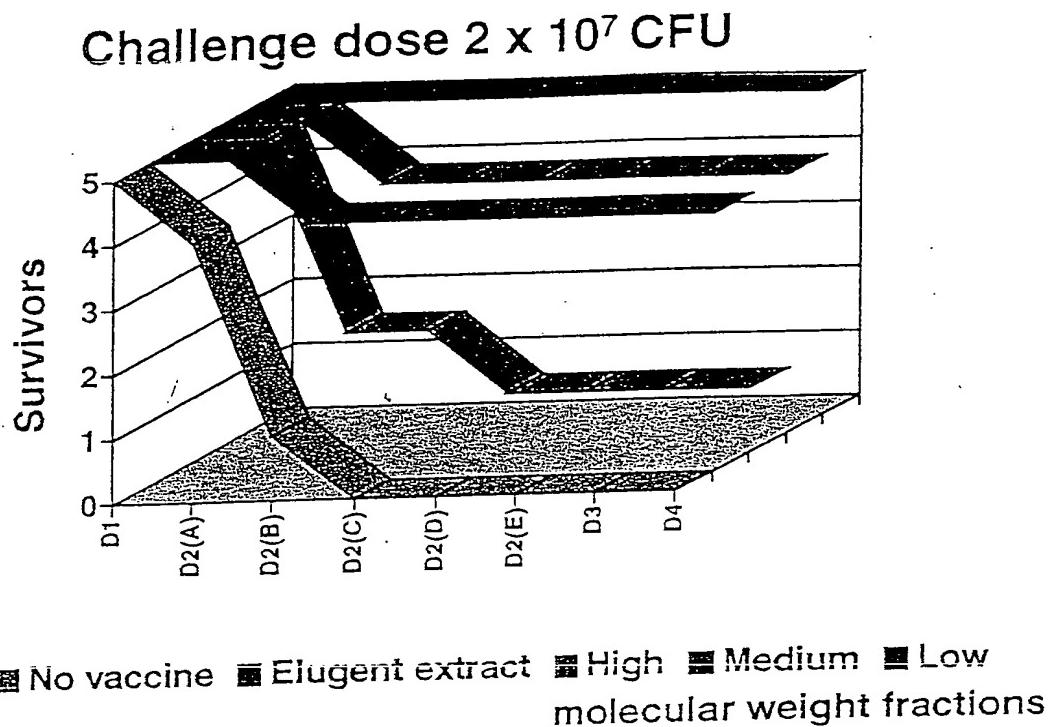
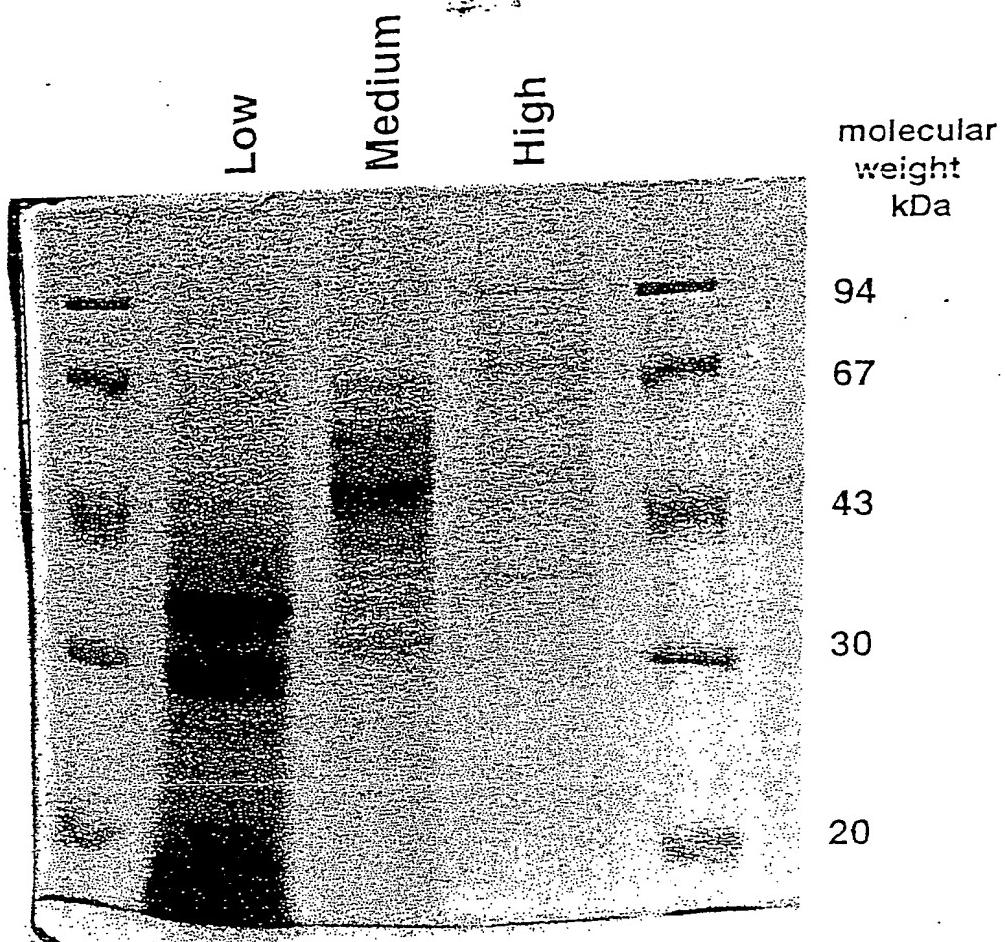


Fig. 2B. SDS-PAGE of proteins separated by preparative electrophoresis used in protection expt.



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Fig 3. Immunoblot showing cross-reaction of antibodies in sera from meningococcal disease patients with proteins from *N. lactamica* strain Y92-1009

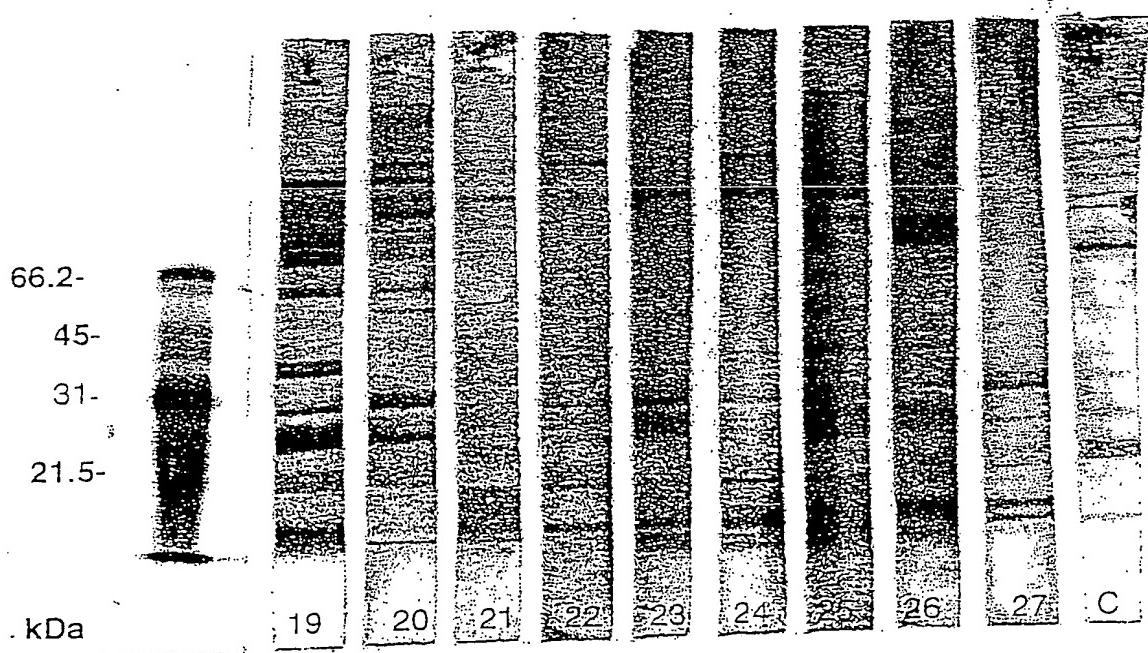


Fig.4

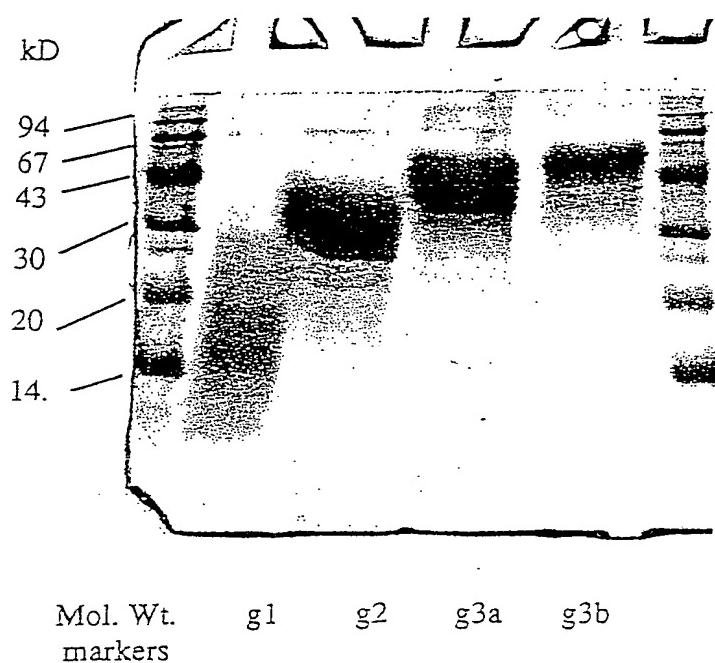
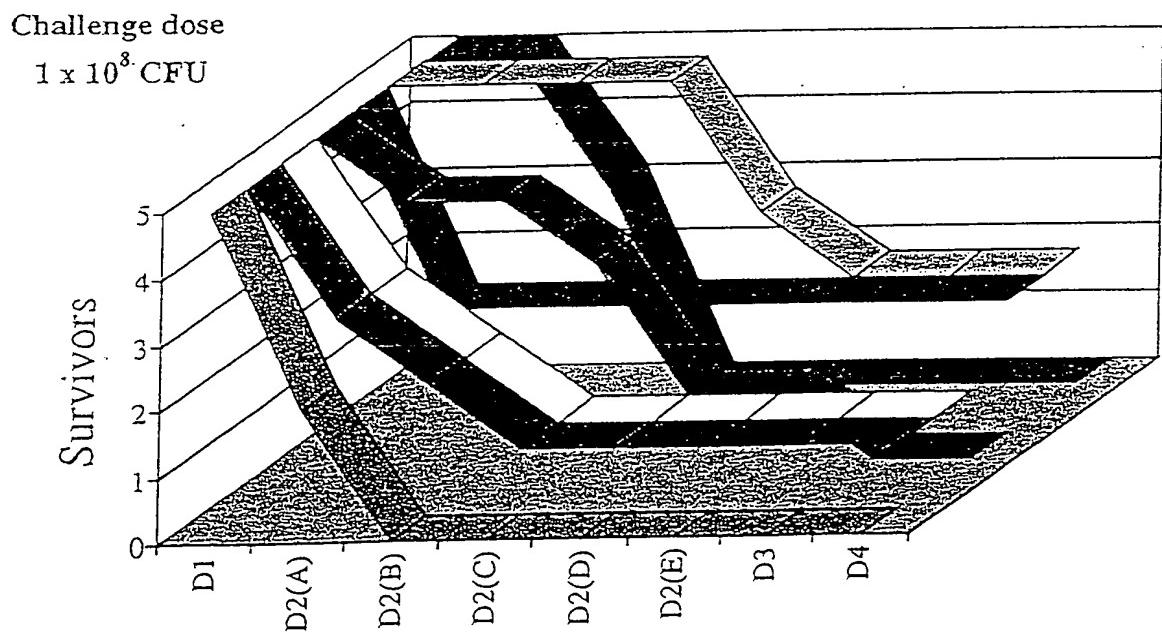
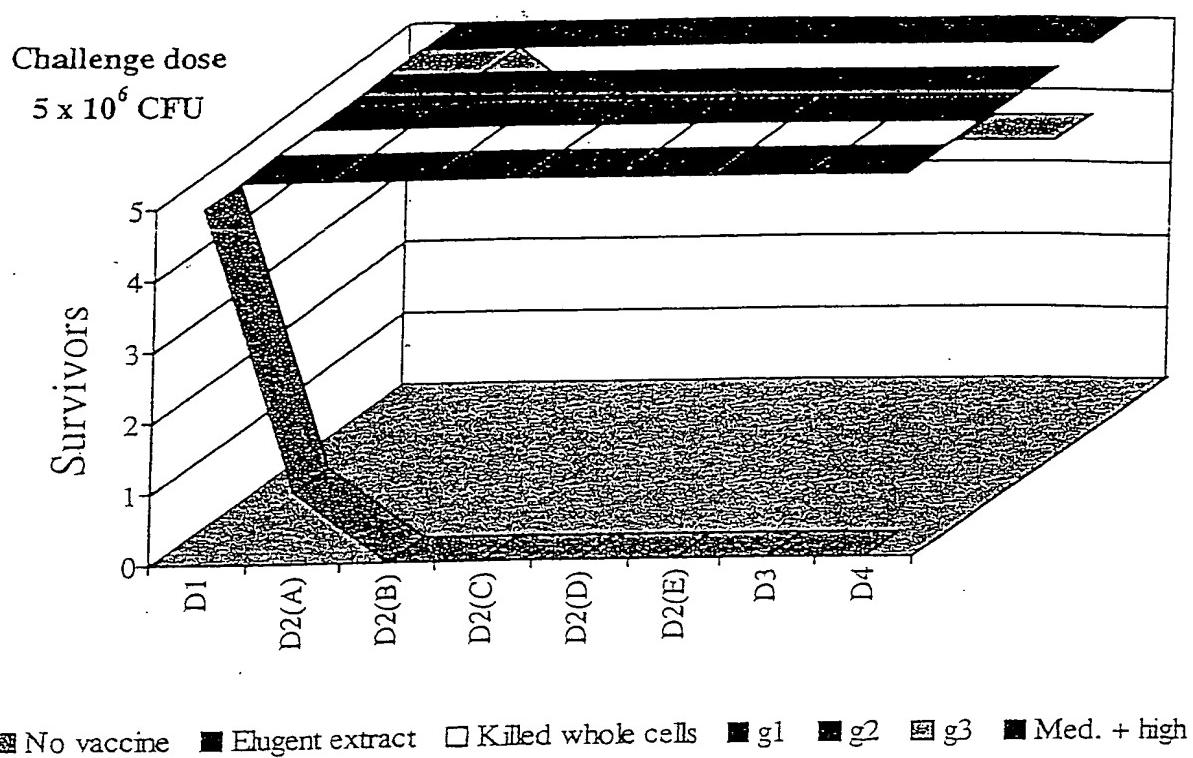


Fig. 5



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Fig. 6

